CatchGene™ Cell/Blood DNA Kit

Cat. No.	Rxn
MD10004	4
MD10050	50
MD10250	250

Kit Content

	4rxn	50rxn	250rxn	
Spin Column	4	50	250	pcs
Collection Tube (2 ml)	12	150	750	pcs
PK Solvent	0.5	1.5	10	ml
Proteinase K	1	11	55	mg
Buffer DL	0.96	12	60	ml
Buffer W1 (concentrated)	3.36	42	105x2	ml
Buffer W2 (concentrated)	0.68	8.4	42	ml
Elution Buffer	0.96	12	60	ml

Important Notice !
"**Proteinase K**" should be stored at -20°C upon arrival for long term storage.

If a precipitate has formed in Buffer DL , dissolve by incubating at 60°C.

Kit Preparation

1. **Prepare 10 mg/ml Proteinase K**
   For 1 mg Proteinase K, please add 100 µl PK Solvent into tube and vortex thoroughly for dissolving.
   For 11 mg Proteinase K, please add 1100 µl PK Solvent into tube and vortex thoroughly for dissolving.
   For 55 mg Proteinase K, please add 5.5 ml PK Solvent into bottle and vortex thoroughly for dissolving.
   After dissolving into solvent, plase store in 4°C for 6 month or -20°C for 1 year.
2. **Prepare Buffer W1**
   Add equal volume of 100% EtOH into Buffer W1 (concentrated) to get Buffer W1.
   After adding 100% EtOH, please check the sticker on the bottle and close the cap tightly.
3. **Prepare Buffer W2**
   Add 4 volume of 100% EtOH into Buffer W2 (concentrated) to get Buffer W2.
   After adding 100% EtOH, please check the sticker on the bottle and close the cap tightly.

Sample Pretreatment

Important! Please store at 4 ℃ if you don't extracts whole blood immediately. Even if you store at 4 ℃ do not over 10 days.

1. The fresh whole blood do not store at room temperature over 2hours.
2. If your whole blood sample were stored at 4°C. Please thaw samples at room temperature (15-25°C) 15 minutes.
3. When you extraction whole blood sample, you need to inverting the blood collection tube 20 times to mix the sample.

General Protocol

1. Add 20 µl Proteinase K into bottom of 1.5 ml micro-centrifuge tube.
2. Add 200 µl whole blood sample into 1.5 ml micro-centrifuge tube, vortex for 5 sec then brief spin down.
3. Add 200 µl Buffer DL, vortex for 15 sec then brief spin down. Incubate at 60 °C for 10 min.
4. Brief centrifuge then add 200 µl 100% EtOH, pluse-vortexing for 15 sec then brief spin down.
5. Place the Spin Column into a new Collection Tube (2 ml).
6. Transfer all mixture to Spin Column, centrifuge at 11,000 x g for 1 min, discard the flow-through and change a new Collection Tube.
7. Add 700 µl Buffer W1 into Spin Column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
8. Add 700 µl Buffer W1 into Spin Column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
9. Add 700 µl Buffer W2 into Spin Column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
10. Change a new Collection Tube, centrifuge at 11,000 x g for 3 min.
11. Place Spin Column into a 1.5 ml micro-centrifuge tube, add 30-200 µl Elution Buffer and incubate at room temperature for 3 min, centrifuge at 11,000 x g for 1 min for elution.

FOR RESEARCH USE ONLY