

Cat. No.	Rxn
MR10004	4
MR10050	50
MR10250	250

## Kit Content

	4rxn	50rxn	250rxn	
Spin Column	4	50	250	pcs
Collection Tubes (2 ml)	12	150	750	pcs
Buffer RC	7.2	90	225x2	ml
Buffer RL	1.68	21	105	ml
Buffer RB	0.5	6.3	31.5	ml
Buffer RW1 (concentrated)	1.68	21	105	ml
Buffer RW2 (concentrated)	0.96	12	60	ml
RNase-Free H <sub>2</sub> O	0.96	12	60	ml

## Kit Storage

Columns, buffers, solvents and consumables, please store at 15-25 °C.

## Kit Preparation

- 1. Prepare Buffer RB**  
Add 2.4 volume of 100% EtOH into Buffer RB and vortex thoroughly.  
After adding 100% EtOH, please tick the sticker on the bottle and close the cap tightly.
- 2. Prepare Buffer RW1**  
Add equal volume of 100% EtOH into Buffer RW1 (concentrated) to get Buffer RW1.  
After adding 100% EtOH, please check the sticker on the bottle and close the cap tightly.
- 3. Prepare Buffer RW2**  
Add 4 volume of 100% EtOH into Buffer RW2 (concentrated) to get Buffer RW2.  
After adding 100% EtOH, please check the sticker on the bottle and close the cap tightly.

## General Protocol

1. Pipette 200 µl fresh whole blood sample into 1.5 ml micro-centrifuge tube and add 1000 µl Buffer RC, mix well by inversion.
2. Incubate on ice for 15 min. (Mix 2 times by inversion during incubation.)
3. Centrifuge at 400 x g for 10 min at 4 °C to form a cell pellet and discard the supernatant completely.
4. Add 400 µl of RC Buffer to res-suspend the cell pellet and mix well by inversion.
5. Centrifuge at 400 x g for 10 min at 4 °C to form a cell pellet and discard the supernatant completely.
6. Add 350 µl Buffer RL (add 1% β-mercaptoethanol freshly), vortex vigorously for 30 sec, brief spin down then incubate at 25 °C (room temperature) for 5 min.
7. Add 350 µl Buffer RB, mix by pipetting and transfer all mixture to Spin Column (in 2ml Collection Tube).
8. Centrifuge at 11,000 x g for 1 min, discard the flow-through and change a new collection tube.
9. Add 700 µl Buffer RW1 into spin column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
10. (Optional) On column digest of DNA with DNase I (not provided).
11. Add 500 µl Buffer RW2 into spin column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
12. Add 500 µl Buffer RW2 into spin column, centrifuge at 11,000 x g for 1 min, discard the flow-through.
13. Change a new collection tube, centrifuge at maximum speed (~18,000 x g) for 3 min.
14. Place the spin column into 1.5 ml micro-centrifuge tube, add 30-100 µl RNase-Free H<sub>2</sub>O, centrifuge at 11,000 x g for 1 min for elution.

**FOR RESEARCH USE ONLY**