Condalab

Reference: 6019 **Technical Data Sheet**

Product: **PRESTON CAMPYLOBACTER SUPPLEMENT**

Specification

Sterile selective supplemet for the isolation *Campylobacter spp.* from human, animal, avian and environmental specimen.

Presentation

	Packaging Details	Shelf Life	Storage
10 Freeze dried vials Vial with: 3 ± 0.1 g	22±0.25 x 55±0.5 mm glass vials, tag labelled, White plastic cap - 10 vials per box.	49 months	2-25 ºC

Composition

Compositon (g/vial)

Polymyxin	2500 IU
Rifampicin	0.005
Trimethoprim	0.005
Cycloheximide	0.050

NOTE : Each vial is sufficient to supplement 500ml of Campylobacter PRESTON medium Base

Reconstitute the original freeze-dried vial by adding
Ethanol..... 6 ml

Description /Technique

Description:

The use of this supplement, added to Campylobacter Medium Base, with 5-7% lysed defibrinated horse or sheep blood, permits the isolation of *Campylobacter* spp. inhibiting the companion flora.

Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.
Reconstitute the vial with the sterile diluent in aseptic conditions and add it to 500 ml of medium base cooled to 50ºC, previously supplemented also with 5-7% lysed defibrinated horse or sheep blood and with Campylobacter Growth Supplement.
Do not overheat once supplemented.
- Agar Base:
Pour the complete medium into Petri dishes and, once solidified on a flat surface, spread the plates either by streaking or by spiral method.
Incubate the medium in microaerophilic conditions at 35 ± 2°C o 42 ± 2°C for 24-48h.
Campylobacter spp. best grown at 42°C.
Incubation times longer than those mentioned above or different incubation temperatures may be requied depending on the sample or the specifications).
After incubation, count all the colonies that have appeared onto the surface of the agar.
-Broth Base:
Dispense the complete medium into suitable containers and inoculate them with the tested specimens.
Incubate the medium in microaerophilic conditions at 35 ± 2°C o 42 ± 2°C for 24-48h.
Campylobacter spp. best grown at 42°C.
Incubation times longer than those mentioned above or different incubation temperatures may be requied depending on the sample or the specifications).
After incubation, subculture on Preston Campylobacter Selective Agar or Campylobacter Blood-Free Selective Agar.
In any case presumptive isolation of *Campylobacter* spp. must be confirmed by further microbiological and biochemical tests.

Quality control

Physical/Chemical control

Color : Orange pH: at 25ºC

Microbiological control

Reconstitute 1 vial as indicated in COMPOSITION; shake and dissolve completely
Add 1 vial to 500 ml of medium base. DO NOT HEAT once supplemented.

Distribute the complete medium, cooled to 50 ºC, into 90 mm plates

Incubate according instructions for complete medium indicated in COMPOSITION.

Microaerofilic incubation at 35 ± 2 ºC or 42± ºC for 24-48 h

Microorganism	Growth
Camp. coli-jejuni ATCC® 33291, WDCM 00005	Good
Campylobacter jejuni ATCC® 29428, WDCM 00156	Good
Escherichia coli ATCC® 25922, WDCM 00013	Inhibited
Stph. aureus ATCC® 25923, WDCM 00034	Inhibited

Sterility Control

Add 5 ml of the sample to:
100 ml TSB and 100 ml Thioglycollate.
Incubation 48 hours at 30-35 ºC and 48 hours at 20-25 ºC: NO GROWTH.

Bibliography

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· CORRY, J.E.L., H.I. ATABAY, S.J. FORSYTHE & L.P. MANSFIELD (2003) Culture Media for the Isolation of Campylobacters, Helicobacters and Arcobacters, en Corry et al. (Eds) Handbook of Culture Media for Food Microbiology Chap 18 pgs 271-316. Elsevier Science B.V. Amsterdam.

· ISO 10272-1 Standard (2017) Microbiology of the food chain - Horizontal Method for detection and enumeration of Campylobacter spp. - Part 1: Detection method.

. ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.