

Urea Broth

For the differentiation of Enterobacteria, particularly Proteus from Salmonella and Shigella from clinical samples.

Categories

Enterobacteria

Enterobacteria

Practical information

Aplications Confirmation Differentiation

Industry: Water / Clinical / Food

Regulations: BAM





Cat. 1226

Principles and uses

Urea Broth can be used for the determination of the urea activity of Enterobacteriaceae, as well as microorganisms of the families of Brucella, Bacillus, Micrococcus, Mycobacteria and Proteus. It can be used for the identification of bacteria on the basis of urea utilization. It is especially recommended for the differentiation of members of the genus Proteus from those of Salmonella and Shigella.

Urea is a source of nitrogen for those organisms producing urease. Yeast extract is a source of vitamins, particularly of the B-group essential for bacterial growth. Potassium phosphates provide buffering capacity. Phenol red is the pH indicator.

When organisms utilize urea, ammonia is produced during incubation, making the reaction of these media alkaline. Positive urease tubes turn the phenol indicator a deep violet-red color (alkalinization). Therefore, urease production may be detected by a change in the phenol red indicator.

Developed by Rustigian and Stuart, this highly buffered medium usually reacts only to the high outputs of ammonia by Proteus, Morganella and Providencia in the first 24 hours of incubation.

Formula in g/L

| Disodium phosphate 9,5 Monopotassium ph | | Monopotassium phosphate | 9,1 |
|---|------|-------------------------|-----|
| Phenol red | 0,01 | Urea | 20 |
| Yeast extract | 0,1 | | |

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 3,87 grams of the medium in 100 ml of distilled water without heating. When the powder is dissolved, sterilize by filtration. Dispense quantities of 0,5 to 2 ml in small sterile tubes. Larger volumes can be used but the reactions will be slower. Do not sterilize in autoclave. Do not boil the medium.

When there is no filter available, the medium can be sterilized at 100 -110°C for 10 minutes. If the medium is prepared and inoculated immediately it provides good results without sterilizing.

Instructions for use

For clinical diagnosis, the type of sample is bacteria isolated from urine and feces:

- Prepare a heavy suspension of the organism isolated from plated media and inoculate the Urea Broth tubes.

- Incubate at 37 °C for 24 hours.



Quality control

| Solubility | Appareance | Color of the dehydrated medium | Color of the prepared medium | Final pH (25°C) |
|------------|-------------|--------------------------------|------------------------------|-----------------|
| w/o rest | Fine powder | Pink | Red-orange | 6,8±0,2 |

Microbiological test

Incubation conditions: (37 °C / 24 h) Inoculation conditions: Confirmation (isolated colony)

| Microorganisms | Characteristic reaction |
|-----------------------------------|--|
| Salmonella enteritidis ATCC 13076 | Urease (-): No liberation of ammonia, no change of colour |
| Salmonella typhimurium ATCC 14028 | Urease (-): No liberation of ammonia, no change of colour |
| Escherichia coli ATCC 25922 | Urease (-): No liberation of ammonia, no change of colour |
| Shigella flexneri ATCC 29903 | Urease (-): No liberation of ammonia, no change of colour |
| Proteus mirabilis ATCC 29906 | Urease (+): Liberation of ammonia with colour change to rose/rose-pink/deep cerise |

Storage

Temp. Min.:2 °C Temp. Max.:8 °C

Bibliography

Rustigian and Stuart. Proc. Soc. Exp. Biol. and Med. 47:109, 1941. Stuart, Van Stratum and Rustigian. J. Bact. 48:437. 1945. McKay, Edwards and Leonar A. J. Clin. Path. 17:479, 1947. Gordon and Mihn. J. Gen. Microbiol., 21:736. 1959. Goldsmith and Latlief. Applied Microbiol., 3:195. 1955.